

## Anigen SYBR green RT-PCR Kit

### Principles

Anigen SYBR green RT-PCR Kit is designed for easy and accurate real-time quantification of RNA from PCR sample. SYBR green I mixed master mix can detect PCR products without target specific probe.

Anigen SYBR green RT-PCR Kit uses Hot Start Taq DNA polymerase, and the RT-PCR buffering conditions are finely adjusted to provide optimum results. ROX reference dye is optional for selection according to Real-Time PCR apparatuses.

The SYBR green I binds to double stranded DNA as a fluorescent dye, and the melting curve analysis is shown specific amplification of the target.

### Materials provided

- 1) 2x SYBR green RT-PCR master mix .....25ml  
: Hot start Taq DNA polymerase, SYBR green Master mix buffer, dNTP mix, SYBR green I, ROX reference dye, MgCl<sub>2</sub>
- 2) RT-mix.....500  $\mu$ l
- 3) ROX reference dye\* .....1ml
- 4) RNase free water.....20ml
- 5) Inset.....1 EA

\* ROX Reference Dye: ROX reference Dye can be included in the reaction to normalize the fluorescent reporter signal, for instruments that are compatible with that option. ROX is supplied at a 25 $\mu$ M concentration, and is composed of a glycine conjugate of 5-Carboxy-rhodamine, succinimidyl ester in 20 mM Tris-HCl (pH 8.4), 0.1mM EDTA, and 0.01% Tween 20. Use the following table to determine the amount of ROX to use with a particular instrument:

Instrument	Amount of ROX per 50 $\mu$ l reaction	Final ROX Concentration
ABI 7000, 7300, 7900HT, and 7900 HT Fast	1 $\mu$ l	500nM
ABI 7500; Stratagene Mx3000 <sup>TM</sup> , MX3005P <sup>TM</sup> , and MX4000 <sup>TM</sup>	0.1 $\mu$ l	50nM

-To accurately pipette 0.1  $\mu$ l per reaction, we recommend that you dilute ROX 1:10 immediately before use and use 1  $\mu$ l of the dilution.

### Storage and Stability

Store at -20 $^{\circ}$ C. 2x SYBR green RT-PCR master mix should be protected from light.

### Procedure

- 1) Thaw each reagent in ice.
- 2) Prepare PCR reaction mixture as below

Components	Volume/reaction	Final concentration
2x SYBR green RT-PCR master mix	25 $\mu$ l	1x
Primer A	Variable	0.5 $\mu$ M
Primer B	Variable	0.5 $\mu$ M
RT-mix	1 $\mu$ l	
Template RNA	Variable	$\leq$ 500ng/reaction
RNase free water	Variable	
ROX reference dye	1 $\mu$ l	
Optional: Uracil-N-glycosylase	Variable	1~2units/reaction
Total reaction volume	50 $\mu$ l	

\* Add the Template RNA at 4 step.

- 3) Mix the PCR reaction mixture completely, and dispense PCR tubes or Plate wells.
- 4) Add the Template RNA to the individual tube or well.

5) Set up the Real-Time PCR cyclers as below.

Step	Time	Tem.	Additional Comments	
Reverse transcription	30min	42 $^{\circ}$ C	cDNA synthesis step	
PCR initial activation step	15min	95 $^{\circ}$ C	Hot Start Taq DNA Polymerase is activated by this heating step	
3-step cycling	Denaturation	15sec	94 $^{\circ}$ C	
	Annealing	30sec	50~60 $^{\circ}$ C	Approximately 5~8 $^{\circ}$ C below T <sub>m</sub> of primers
	Extension	30sec	72 $^{\circ}$ C	Perform fluorescence data collection, unless an additional data acquisition step has been intergraded
	Number of cycle	35~45cycle		Number of cycles depends on the amount of template RNA and transcript abundance

- 6) Place the PCR tubes or Plate in the Real-time PCR cycler and start the program.
- 7) Perform the melting curve analysis.

8) Optional: Confirm the amplification using electrophoresis for Specificity of Real-Time PCR.